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Chromatographic fractionation, antioxidant and antibacterial activities of *Urginea maritima* methanolic extract

Oum Elkheir Belhaddad¹, Noureddine Charef¹, Samra Amamra¹, Fatima Zerargui¹, Abderrahmane Baghiani¹, Seddik Khennouf² and Lekhmici Arrar¹*

¹Laboratory of Applied Biochemistry, Faculty of Nature and Life Sciences, University Setif, Setif 1, Algeria

²Laboratory of Phytotherapy Applied to Chronic Diseases, Faculty of Nature and Life Sciences, University Setif 1, Setif, Algeria

Abstract: The present work concerns a phytochemical study of *Urginea maritima* L. from Algeria, and an evaluation of antioxidant activity of the methanolic extract (UMME) and its chromatographic fractions. UMME was fractionated using open glass chromatography on silica gel and antioxidant effects were evaluated using DPPH and β -carotene/linoleate assays. The phytochemical screening revealed that the bulb of plant contains flavonoids, glycosides, tannins, reducing compounds, anthraquinones combined, anthocyanins, mucilage, triterpenes and steroids. DPPH method showed that the UMME has a scavenger effect on radical DPPH with an IC₅₀=57.83±1.59µg/ml. The fractions isolated from *U. maritima* (L.) presented an IC₅₀ ranging between 499.23 and 39.68µg/ml. In of β -carotene/linoleate test, UMME and fractions give an I% =69.56±0.08% and between 31.29±0.49% and 90.79±0.29%, respectively. UMME showed a high inhibitory effect on the xanthine oxidase (IC₅₀=0.67±0.01 mg/ml) and on the cytochrome c reduction (IC₅₀=0.68 mg/ml). Wide range of phytochemical constituents in *Urginea maritima* were detected in methanolic extract which exhibited antioxidant and antibacterial activity. This plant could serve as pilot for the development of novel agents for pathological disorders.

Keywords: Urginea maritima, phytochemical screening, column chromatography, Antioxidant, DPPH, secondary metabolites.

INTRODUCTION

Urginea maritima L. Baker is a perennial bulbous geophyte (a herbaceous plant with an underground storage organ) of the family Liliaceae (Bruneton 1996), native to the Mediterranean basin and well-adapted to its type of climate (Kopp et al., 1996). It generally occurs in the slopes of hills, the sandy grounds near the Mediterranean Sea and in certain regions of Northern Africa (Bellakhdar, 1997), Middle East and Europe. Urginea maritima has two varieties: Red and white. The red variety (red squill) is predominant in Algeria (Sandberg and Corrigan, 2001) and Greece (Altardeh et al., 2006). The white variety is predominant in Morocco (Bellakhdar, 1997). From the phytochemical point of view, it has been reported that the major constituents of U. maritima bulbs are glycosides (Kopp et al., 1996; Krenn et al., 2000), Anthocyanins (Vega et al., 1972), flavonoids (Fernandez et al., 1972), fatty acids, polysaccharides (Spies et al., 1992) and calcium oxalate (Cogne et al., 2001). The cardiac glycosides (scillaren and scillarenin) are used in Europe as a cardiotonic diuretic for the treatment of cardiac marasmus and edema (Mitsuhashi et al., 1994). It was then expected that this plant inhibited Na⁺/K⁺ adenosine triphosphatase (Schonfeld et al., 1985). Furthermore, Bayazit and Konar (2010) showed that Squill bulb scillioside can reduce the musculoskeletal pains. It has been shown that extracts of bulbs have been active against stored product pests. This

suggests that squill should be investigated for activity against other insects and pests (Pascual-Villalobos and Fernández, 1999). The objective of this research was to study the phytochemical and the fractionation of the methanolic extract of U. maritima. The evaluation of antioxidant activity and antibacterial effects of the methanolic extract and each chromatographic fraction was also carried out.

MATERIALS AND METHODS

Reagents

All reagents used were of analytical grade. Tween-20, methanol, ethanol, benzene, acetone, β -carotene, chloroform, linoleic acid were purchased from E. Merck (Darmstadt, Germany). 1,1-diphenyl-2-picrylhydrazyl (DPPH), butylated-hydroxytoluene (BHT) were obtained from sigma Chemical Co. (St. Louis, MO). All other chemicals and solvents were analytical grade.

Plant material

Urginea maritima bulbs were collected at Bordj Bou Arreridj, Algeria, from September to October 2010 and were identified by Prof H. Laouer, Department of Ecology and Vegetal Biology, Faculty of Nature and Life Sciences, University Setif. Bulbs parts of the plant were air dried for several days. The dried plant material was ground to a coarse powder using a dry mill. For the extraction procedure, 1kg of powdered plant material was soaked in methanol for 7 than 5 days at room temperature with renewal of the solvent (Markham, 1982). The solvent

^{*}Corresponding author: e-mail: lekharrar@hotmail.com

was then removed by rotary evaporation to obtain a dried extract called *Urginea maritima* methanolic extract (UMME).

Phytochemical screening

The screening is important to the next steps like extraction and/or fractionation of extracts to isolate compounds of interest. The phytochemical tests enable to detect the presence different groups of biochemical substances. Reducing compounds and alkaloids were performed according to the method described by Sofowora (1985) and Bruneton (2009). The tests were based on the extraction with suitable solvents of increasing polarity and the visual observation of color change or formation of a precipitate after the addition of specific reagents. These reactions are selective to types or groups of compounds, simple, rapid, sensitive and require a minimum of laboratory equipment.

Fractionation of methanolic extract

Open glass column chromatography, 100cm high and 4 cm of diameter, of UMME was carried out on silica gel 70-230 mesh, packed in dichloromethane. Elution was carried out using dichloromethane-methanol of increasing polarity. After TLC analysis, similar fractions were combined into pools called here fractions. The pool (fraction) 26 was subjected to further purification on a little column of 30cm high and 2cm of diameter using solvent mixtures of chloroform-methanol-acetic acid-H₂O of increasing polarity (Abbot and Andrews, 1970).

HPLC analysis

HPLC analysis was performed using a HPLC Model 9100 apparatus equipped with a ternary pump Model 9100 and UV–visible detector Model 9100. A reverse phase column C18 (250 x 4.6mm, 5 μ m particles) was used. The mobile phase consisted of a binary mixture of methanol and water (60:40 v/v) at isocratic flow rate of 1ml. min⁻¹. The absorbance was monitored at λ =254 (Kuntić *et al.*, 2007).

Determination of total Polyphenol and flavonoids Contents

The total polyphenols in *Urginea maritima* extracts were determined by the Folin–Ciocalteu method according to Cliffe *et al.* (1994) with slight modifications as we have previously described (Baghiani *et al.*, 2012). Flavonoids were quantified using aluminium chloride reagent (AlCl₃), (Bahorun *et al.*, 1996). They were measured as quercetin and rutin equivalents. One ml of *Urginea maritima* extracts was dissolved in methanol, 1ml of AlCl₃ (2 %) in methanol was added, after incubation for 10 min, the absorbance was measured at 430 nm.

Free radical scavenging activity using DPPH

The free radical scavenging properties of Urginea maritima extracts were measured by decrease in the

absorbance of methanol solution of DPPH (Burits and Bucar, 2000; Baghiani *et al.*, 2012).

Antioxidant assay using β -carotene-linoleate test

 β -carotene bleaching assay was carried out according to the method described by Dapkevicius *et al.* (1998) with modifications (Baghiani *et al.*, 2012). In this test, BHT was used as positive control and MeOH and H2O as blanks.

Effects of methanolic extract of Urginea maritima on xanthine oxidase activity

The effect of extract methanolic of *Urginea maritima* on xanthine oxidase (XO) activity was determined by measuring the absorbance 295 nm (Boumerfeg *et al.*, 2009). The enzymatic reaction is initiated by the addition of bovine XO, which was prepared in our laboratory with a specific activity of 1176 nmole/min/mg of enzyme, to the xanthine prepared in phosphate buffer containing EDTA as we have previously described (Boumerfeg *et al.*, 2009). Allopurinol was used as a positive standard. The effects on the superoxide anions generation by XO was measured by following the cytochrome c (25 mM) reduction at 550 nm (Robak and Gryglewski, 1988).

Antimicrobial activity

The antibacterial activity of UMME was tested against 10 bacteria strains: Salmonella typhimurium ATCC 13311, Acinetobacter baumanii ATCC 19606, Klebsiella pneumoniae ATCC 700603, Bacillus cereus ATCC 10876, Enterobacter cloaceae, Listeria monocytogenes ATCC 15313, Escherichia coli ATCC 25922, Pseudomonas aeruginosa ATCC 27853, Staphylococcus aureus, ATCC 25923 and Proteus vulgaris. The disc difusion technique was used to evaluate the antibacterial activity of methanolic extracts. Ten µl of powder resin by disc of Whatman paper N°1 was applied into Muller Hinten medium inoculated by a bacterial suspension. The bacterial solution was diluted in order to obtain a concentration equivalent to 0.5Mc Farland (NCCLS, 2001). The dried material of plant extracts was dissolved at different concentrations in one ml of DMSO. Bacterial suspensions (100µl) were spread on Muller Hinton Agar medium. The extracts (10µl) were applied to discs of Whatman paper N°1 and applied into the medium inoculated by a bacterial suspension. The plates were incubated at 37°C for 48h.

RESULTS

Phytochemical screening

Urginea maritima powder was subjected to various phytochemical tests to identify the present chemical constituents. The results showed the presence of flavonoids, glycosides, tannins, reducing compounds, anthraquinones combined, anthocyanins, mucilage, triterpenes and steroids (table 1).

Table 1: Result of phytochemical screening of Urginea maritima

| | Test | Results | |
|--------------------------|-----------|---------------|-------|
| Alkaloids | | - | |
| T | Catechols | | +++ |
| Tannins | Gallic | | + |
| Anthocyanins | | | ++ |
| Flavonoids | | | ++ |
| Leucoanthocyans | | | +++ |
| | | Free | - |
| Anthraquinones | combined | o-heterosides | + |
| | combined | c-heterosides | + |
| triterpenes and steroids | | | + + + |
| Saponins | | | - |
| Reducing compounds | | | + |
| Glycosides | | | + + + |
| Coumarins | | | - |
| Mucilage | | | + + |

+ : present, - : absent

| | | | | | | Meth | nanolio | extra | act of | Urgine | ea mar | itima | | | | | | |
|-------|------------|--------|--------|---------------|---------|---------|---------|--------|-----------------|--------------------|---------|-------------------|---------|---------|---------|---------|---------|-------|
| | | | | | | | | | CH ₂ | Cl ₂ /M | eOH/C | H ₃ CO | OH | | | | | |
| Pools | 5 1 | 2 | 3 | ↓ 4 | 5 | 6 | 7 | 8 | 9 | 10 | ↓ 11 | 12 | 13 | 14 | 15 | 16 | ↓ 17 | 18 |
| Fract | ions 1-5 | 6-9 | 10 | 11-14 | 15-18 | 19-21 | 22-23 | 24-25 | 26 | 27 | 28 | 29 | 30 | 31-35 | 36-37 | 38 | 39 | 40-43 |
| Mixtu | re 100/0/0 | 99/1/0 | 98/2/0 | 97/3 /0 | 95/5 /0 | 90/10/0 | 88/12/0 | 85/1/0 | 83/17/0 | 83/17/0 | 80/20/0 | 75/25/0 | 70/30/0 | 70/29/1 | 70/28/2 | 70/25/4 | 70/25/5 | MeOH |
| Mass | 6 (mg) 810 | 470 | 90 | 430 | 770 | 1180 | 620 | 1470 | 4870 | 2280 | 2260 | 780 | 6830 | 3480 | 3030 | 4900 | 3600 | 1404 |
| | | | | | | | | | С | HCI ₃ / | MeOH | CH ₃ C | OOH | | | | | |
| | Fractions | Į | 8 | 2 | 3 | 4 | ţ | | 6 | 1 | 8 | 9 | 10 | 11 | | 12 | | |
| | Mixture | 100/0 |)/0 9 | 8/2/0 | 95/5/0 | 94/6/0 | 92/8/ | | | , 35/15/0 | 82/18/0 | 78/22/0 | | | | 40/10 | | |
| | Mass (mg |) 20 | | 520 | 220 | 130 | 170 | 1 | 340 | 900 | 150 | 80 | 50 | 20 |) | 40 | | |

Fig. 1: The fractionation procedure of Urginea maritima methanolic extract by column chromatography in two steps.

Fractionation of methanolic extract

A total of 43 fractions were collected. These were pooled, based on similar thin-layer chromatograms into 18 pools. Results of column chromatography of the methanolic extract are shown in fig. 1. The bioactive fraction F26 (4.87g) was subjected for further fractionation (purification) on column chromatography. Elution was done with chloroform and chloroform-methanol to get 12 pooled sub fractions.

The nature of the sample source determines the isolation techniques, which vary among solid and liquid samples. Polyphenols and flavonoids in plant were isolated by extraction with chloroform-methanol, followed by separation on column chromatography. Elution was done with chloroform and chloroform-methanol, for obtaining enriched total flavonoids and polyphenols.

Total polyphenols and flavonoids contents

Table 2 presents the amount of phenolic and flavonoïds compounds in methanolic extracts of *U. maritima* and its chromatographic fractions. Total phenolic contents were expressed as mg Gallic acid equivalents per gram of dry weight (mg GA Eq/g extract) and total flavonoids contents as mg Quercetin and Rutin equivalents per gram of dry weight (mg Q Eq and R Eq/g extract).

In UMME fractions, the fraction 25 presents the highest amount of phenolic compounds (404.19 ± 1.52 mg GA Eq/g) followed by crud UMME (table 2). Fractions 1 to 19 contained very low amount of polyphenols and flavonoids. In the fractions of F26, the fraction F11 contained the highest concentration of polyphenols (318.09 ± 0.44 mg GA Eq/g) and total flavonoids contents (32.12 ± 0.4 mg Q Eq/g), (fig. 2).



Fig. 2: Total phenolic and flavonoid contents of fraction 26 and its major chromatographic fractions. Polyphenols as mg GA-Eq/g. (A), Flavonoids as mg Q-Eq/g (B) and as mg R-Eq/g.

Free radical scavenging activity using DPPH method

Natural products isolated from medicinal plants have attracted considerable attention in recent years due to their various pharmacological properties. In the present study, DPPH radicals were used in the test to investigate the scavenging effects of methanolic extract and chromatographic fractions. The results showed that UMME possesses a scavenging effect on DPPH radical $(EC_{50}=57.83\pm1.59\mu g/ml)$. The presence of compounds such as tannins, flavonoids, Anthocyanins and phenols in Urginea maritima extract may be the cause of this effect. The chromatographic fractions investigated showed different levels of DPPH free radical scavenging activity. In fractions of UMME, IC₅₀ values ranged from $39.68\pm0.69\mu$ g/ml to $499.23\pm1.25\mu$ g/ml. The IC₅₀ values of fractions of F26 were ranged between 39.91±0.08µg / ml and $619.76 \pm 4.85 \mu g/ml$ (fig. 3).



Fig. 3: IC_{50} values of plant extracts for free radical scavenging activity by DPPH method. Lower IC_{50} value indicates higher antioxidant activity. (A): Fractions of crud extract, (B): Fractions from faction 26. Each value is represented as mean of three determinations \pm S.D.

Antioxidant assay using β -carotene-linoleate

The antioxidant activities of UMME and its fractions compared to butylated hydroxytoluene (BHT) are presented in fig. 4. The I% value of UMME was 69.65%. The best activity was shown by fraction 38 of methanolic extract with an I % value of 87%. The lowest inhibition was 51.19% shown by the fraction 23. In the fractions of F26, the fraction 12 gave a highest inhibition compared to BHT (fig. 4) with an 1% =90.79%.



Fig. 4: Antioxidant activity of the methanolic extract and its chromatographic fractions, compared to BHT (positive blank) and, H₂O and Methanol (negative blanks) using β -carotene-linoleate assay after 24 hours of incubation. Results are means of three different experiments \pm S.D.

Qualitative HPLC analysis

Due to the complexity of the natural mixtures of phenolic compounds of various plants, it is rather difficult to elucidate their structure and assess the antioxidant and biological potentials. Hence, it was aimed in this work to isolate and identify some of the phenolic compounds present in the methanolic extract and major fractions using HPLC which is a high-resolution chromatographic technique probably the most widely used analytical technique for characterizing the polyphenolic compound (Gomez-Caravaca et al., 2006). Crude extract and four more active fractions of Urginea maritima bulb (F26, F30, F36 and F42) were selected in this analysis. The preliminary results presented in table 3 showed the presence of at ten major compounds in the crude methanolic extract and two to four compounds in the chromatographic fractions.

Effects of Urginea maritima methanolic extract on XO activity

The extract inhibited the activity of xanthine oxidase in a concentration dependent manner (fig. 5) with an IC_{50} (mg/ml) value of 0.67±0.01. Additionally, UMME was able to reduce the cytochrome c^{+3} in concentration-dependent manner. It has a potent scavenging activity of superoxide anion radical with an IC_{50} value of 0.68± 0.001mg/ml (fig. 5).

| Extracts | Total polyphenols (mg GA-Eq/g) | Total flavonoids | | |
|----------------|--------------------------------|------------------|-------------|--|
| | Γ | (mg Q-Eq/g) | (mg R-Eq/g) | |
| L'extrait brut | 234.25±0.35 | 11.42±0.35 | 18.69±0.57 | |
| Fraction 20 | 32.38±0.05 | $1.84{\pm}0.02$ | 2.88±0.11 | |
| Fraction 21 | 101.07±0.3 | 5.14±0.05 | 8.37±0.15 | |
| Fraction 22 | 178.3±0.3 | 7.09±0.43 | 11.49±0.74 | |
| Fraction 23 | 88.23±0.23 | 6.48±0.53 | 10.58±0.89 | |
| Fraction 24 | 235.23±0.26 | 17.92±0.62 | 29.48±1.08 | |
| Fraction 25 | 404.19±1,52 | 25.73±1.88 | 42.35±3.8 | |
| Fraction 26 | 124.52±0.33 | 6.6±0.11 | 10.45±0.42 | |
| Fraction 27 | 103.85±2.62 | 7.41±0.41 | 12.18±0.72 | |
| Fraction 28 | 118.33±1.14 | 8.89±0.38 | 14.58±0.77 | |
| Fraction 29 | 76.71±1.95 | 7.29±0.31 | 11.83±0.45 | |
| Fraction 30 | 196.14±0.2 | 14.97±0.99 | 24.77±0.48 | |
| Fraction 31 | 126.14±1.31 | 15.05±0.33 | 24.65±0.48 | |
| Fraction 32 | 182.42±1.71 | 4.91±0.33 | 7.75±0.55 | |
| Fraction 33 | 181.96±0.45 | 7.53±1.32 | 12.36±2.1 | |
| Fraction 34 | 100±0.2 | 16.58±0.67 | 27.31±1.05 | |
| Fraction 35 | 138.11±0.84 | 9.41±0.13 | 15.45±0.14 | |
| Fraction 36 | 178.42±0.8 | 15.09±0.89 | 20.82±1.02 | |
| Fraction 37 | 177.02±0.03 | 9.35±0.12 | 15.71±0.15 | |
| Fraction 38 | 126.61±0.26 | 20.52±1.78 | 34.83±2.89 | |
| Fraction 39 | 83.61±0.13 | 13.14±0.26 | 23.27±0.5 | |
| Fraction 40 | 123.96±0.55 | 19.89±2.25 | 32.78±3.67 | |
| Fraction 41 | 221,07±1,56 | 13.60±0.24 | 22.37±0.34 | |
| Fraction 42 | 301.71±0.4 | 19.41±1.79 | 31.65±2.89 | |
| Fraction 43 | 92.78±0.3 | 3.48±0.05 | 5.66±0.12 | |

Table 2: Total phenolic and flavonoid contents of *U. maritima* methanolic extract and its major chromatographic fractions.

Table 3: Major compounds identified by HPLC analysis of crude extract (CE) and chromatographic more active fractions of *Urginea maritima* bulb.

| Major Compounds | CE | F26 | F30 | F36 | F42 |
|-----------------|-----|-----|-----|-----|-----|
| Tannic acid | +/- | | | + | + |
| Quercetin | + | | + | | |
| Gallic acid | + | + | | | |
| Reserpin | +/- | | | + | |
| Catechin | + | | | | |
| Caffeic acid | + | + | | | +/- |
| Rutin | +/- | + | + | | |
| Vanilin | + | | | | |
| Kaemferol | + | | | | |
| Naringin | + | + | + | | |

Xanthine oxidase mechanism leads to the production of ROS (Harrison, 2002). These ROS have great effects on different biomolecules (Castro and Freeman, 2001).

Antimicrobial activity

Preliminary screening of the *in vitro* antibacterial activity of methanolic extract was studied against various pathogens. The results showed different antimicrobial properties of plant extracts (table 4). *Bacillus cereus* ATCC10876 was found to be the most inhibited pathogen by the UMME with a diameter of inhibition zone of 11

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mm; followed by *Acinetobacter baumanii* (10mm) and *Salmonella typhimurium* (9mm). Ivancheva *et al.* (2006) attributed this activity to the presence of, secoiridoid glucosides, phenylethanoids and flavonoids contained in the extract.

DISCUSSION

The medicinal plants are rich in secondary metabolites including polyphenols, alkaloids, glycosides and flavonoids. They are of great clinical importance and are widely used in pharmaceutical industry. Polyphenols are known as antioxidant and scavenging compounds against free radicals associated with oxidative stress (Fergusion *et al.*, 2006).



Fig. 5: Inhibition of bovine xanthine oxidase activity (A) and super oxide anion radical generation from xanthine/xanthine oxidase system (B) by the methanolic extract of *Urginea maritima*. Results are expressed as percentage of control where no inhibitor was added. Each value is represented as mean \pm S.D (n=3).

The presence of these compounds such as tannins, flavonoids, proanthocyanidins and phenols in Urginea maritima extract may give credibility to its local use for the management of oxidant related pains. Tannins have been found to have antiviral, antibacterial, anti-parasitic effects, anti-inflammatory, antiulcer and antioxidant property for possible therapeutic applications (Ly et al., 2004; Akiyama et al., 2001). The composition of tannins as observed in this study may justify its traditional usage come anti-inflammatory. Flavonoids, the major group of phenolic compounds are reported for their antimicrobial, antiviral and spasmolytic activity. Flavonoids are able to scavenge hydroxyl radicals, super oxide anion radicals and lipid peroxy radicals, it has been confirmed that pharmacological effect of flavonoids is correlating with their antioxidant activities (Boumerfeg et al., 2009). In the present study we observed the presence of glycosides in the methanolic extract of Urginea maritime. Glycosides have a history of pharmacological effects for their cardiotonic and diuretic effects. Therefore, the concentration of these compounds could contribute synergistically to the significant cardiotonic potency of this plant and thus may support the local usage for the treatment of cardiac marasmus and edema (Mitsuhashi et al., 1994). These results reveal that the plant has quite a number of chemical constituents, which may be responsible of the many pharmacological actions.

Several techniques were used in this study to assess the free radical scavenging and reducing properties of the

methanolic extracts of *Urginea maritima*, along with evaluation of the total flavonoids and polyphenols contents. *Urginea maritima* extract acts as reducing agent, exhibiting antioxidant role in inhibiting and scavenging free radicals and providing protection to humans against infections and degenerative diseases (Nagler *et al.*, 2006). The present study showed the ability of the methanolic extract and its fractions to inhibit hydroxyl radical, with a concentration dependent manner.

Flavonoids and other phenolic compounds of plant origin have been reported as scavengers and inhibitors of lipid peroxidation (Adithya *et al.*, 2013). In fact, the antioxidant activity was dependent upon the presence and quantity of polyphenols and/or flavonoids (Ibraheem *et al.*, 2014). Thus, the degradation rate of β -carotenelinoleate depends on the antioxidant activity of the extracts (Maisarah *et al.*, 2013).

CONCLUSION

The main classes of secondary metabolites such as alkaloids, flavonoids, tannins, sterols and terpenoids, were found in U. maritima. The phytochemical analyses showed that the methanolic extract and chromatographic fractions were rich in polyphenols and flavonoids. Additionally, UMME and some of its fractions exhibited substantial potency in scavenging DPPH radical, inhibiting lipid peroxidation, XO activity and cytochrome c reduction. Finally, methanolic extract has an antimicrobial activity against seven species of bacteria. In conclusion, Urginea maritima methanolic extract contains several constituents which have antioxidant and antibacterial activities. Further studies regarding isolation and purification of active phyto-constituents with broad spectrum of antioxidant and antibacterial activities are under investigation.

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REFERENCES

- Abbot D and Andrews RS (1970). An Introduction to chromatography. 2nd ed. Longman Press, London., pp.72-78.
- Adithya ES, Lakshmi MS and Hephzibah P (2013). In vitro antioxidant, anti-lipid peroxidation activities and HPLC analysis of methanol extracts from bark and stem of Mahonia leschenaultia takeda. *Asian J. Plant. Sci. Res.*, **3**(2): 116-126

- Akiyama H, Fujii K, Yamasaki O, Oono T and Iwatsuki K (2001). Antibacterial action of several tannins against *Staphylococcus aureus*. J. Antimicrob. Chemother., **48**(4): 487-491.
- Altardeh S, Sawidis T, Diannelidi BE and Delivopoulos S (2006). Anatomical studies on the adventitious roots of the geophyte *Urginea maritima* (L.) *Baker. J. Biol. Res.*, **5**: 61-70.
- Baghiani A, Ameni D, Boumerfeg S, Adjadj M, Djarmouni M, Charef N, Khennouf S and Arrar L (2012). Studies of Antioxidants and Xanthine Oxidase Inhibitory Potentials of Root and Aerial Parts of Medicinal Plant *Capparis Spinosa* L. Am. J. Med. Medical Sci., 2(1): 25-32.
- Bahorun T, Gressier B, Trotin F, Brunete C, Dine T, Asseur J, Gazin JC, Pinkas M, Luyckx M and Gazin M (1996). Oxygen species scavenging activity of phenolic extract from Hawthorn fresh plant organs and pharmaceutical preparation. *Arzneim. Forsch.*, **46**(11): 1086-1089.
- Bayazit V and Konar V (2010). Analgesic effects of scilliroside, proscillaridin-A and taxifolin from squill bulb (urginea maritima) on pains. *Digest. J. Nanomat. Biostr.* 5(2): 457-465.
- Bellakhdar J (1997). Traditional pharmacopeia of Morocco (in French). Ibis Press, Paris.
- Boumerfeg S, Baghiani A, Messaoudi D, Khennouf S and Arrar L (2009). Antioxidant properties and Xanthine Oxidase Inhibitory Effects of Tamus communis L. Root Extracts. *Phytother. Res.*, **23**: 283-288.
- Bruneton J (1996). Poisonous Plants Plants dangerous to humans and animals (in French). Tec & Doc Lavoisier, Paris, p.529.
- Bruneton J (2009). Pharmacognosy, phytochemistry: Medicinal plants (in French). 4th edition. Lavoisier. Paris.
- Burits M and Bucar F (2000) Antioxidant activity of *Nigella sativa* essential oil. *Phytother. Res.*, **14**: 323-328.
- Castro L and Freeman BA (2001). Reactive oxygen species in human health and disease. *Nutrition*, **17**(2): 163-165.
- Cliffe S, Fawer MS, Maier G, Takata K and Ritter G (1994). Enzyme assays for the phenolic content of natural juices. *J. Agric. Food. Chem.*, **42**(8): 1824-1828.
- Cogne AL, Marston A, Mavi S and Hostettmann K (2001) Study of two plants used in traditional medicine in Zimbabwe for skin problems and rheumatism: *Dioscora sylvatica* and *Urginea altissima*. J. *Ethnopharmacol.*, **75**: 51-53.
- Dapkevicius A, Venskutonis R, van Beek TA and Linssen PH (1998). Antioxidant activity of extracts obtained by different isolation procedures from some aromatic herbs grown in Lithuania. *J. Sci. Food Agric.*, **77**: 140-146.

- Fergusion L R, Philpott M and Karunasinghe N (2006). Oxidative DNA damage and repair: significance and biomarkers. J. Nutr., **136**(10): 2687S-2689S.
- Fernandez M, Vega FA, Arrupe T and Renedo J (1972). Monocotyledonae liliaceae flavonoids of squill, *Urginea maritima*. *Phytochem.*, **11**: 1534.
- Gómez-Caravaca AM, Gómez-Romero M, Arráez-Román D, Segura Carretero A and Fernández-Gutiérrez A (2006). Advances in the analysis of phenolic compounds in products derived from bees. *J. Pharmaceut. Biomed. Anal.*, **41**: 1220-1234.
- Harrison R (2002). Structure and function of xanthine oxidoreductase: where are we now? *Free radic. Biol. Med.*, **33**: 774-797.
- Ibraheem ZO, Satar M, Abdullah NA, Rathore H, Tan YC, Uldin F, Basri R, Abdullah MH and Edward John E (2014). Antioxidant and cardio protective effect of palm oil leaves extract (standardized ethanolic fraction) in rats' model of saturated fats induced metabolic disorders. *Pak. J. Pharm. Sci.*, 27(1): 1-9.
- Ivancheva S, Nikolova M and Tsvetkova R (2006). Pharmacological activities and biologically active compounds of Bulgarian medicinal plants. Chap. 4 *In*: Phytochemistry: Advances in Research. Editor: Filippo Imperato. Research Signpost 37/661 (2), Fort P.O., Trivandrum-695 023, Kerala, India. pp.87-103.
- Kopp B, Krenn L, Draxler M, Hoyer A, Terkola R, Vallaster P and Robien W (1996). Bufadienolides from Urginea maritima from Egypt. Phytochemistry, 42: 513-522.
- Krenn L, Jelovina M and Kopp B (2000). New bufadienolides from *Urginea maritima* sensu strictu. *Fitoterapia.*, **71**: 126-129.
- Kuntie V, Pejie N, Ivkovie B, Vugie Z, Ilie K, Mieie S and Vukojevie V (2007). Isocratic R-P-HPLC method for rutin determination in solid oral dosage forms. *J. Pharmaceut. Biomed. Anal.*, **43**: 718-721.
- Ly L, Liu SW, Jiang SB and Wu SG (2004) Tannin inhibits HIV-1 entry by targeting gp41. *Acta Pharmacol. Sin.*, **25**(2): 213-218.
- Maisarah AM, Nurul Amira B, Asmah R and Fauziah O (2013). Antioxidant analysis of different parts of Carica papaya. *Inter. Food Res. J.*, **20**(3): 1043-1048.
- Markham KR (1982). Techniques of Flavonoid Identification. Academic Press, London. p.133.
- Mitsuhashi H, Tanaka O, Nozoe S and Nagai M (1994) Chemistry of Organic Natural Products. 4th edition. Nankoudou Press, Tokyo. pp.168-174.
- Nagler R, Reznick A, Shafir Y and Shehadeh N (2006). Free radical related effects and antioxidants in saliva and serum of adolescents with type I diabetes mellitus. *Arch. Oral Biol.*, **40**: 156.
- NCCLS (National Committee for Clinical Laboratory Standards). Performance Standards for Anti–Microbial Susceptibility Testing: Eleventh Informational Supplement. Wayne, PA, USA. NCCLS. Document M100-S11. 2001.

- Pascual-Villalobos MJ and Fernández M (1999). Insecticidal activity of ethanolic extracts of Urginea maritima (L.) Baker bulbs. Ind. Crops Prod., 10: 115-120.
- Robak J and Gryglewski RJ (1988). Flavonoids are scavengers of superoxide anions. *Biochem. Pharmacol.*, 37: 837-841.
- Sandberg F and Corrigan D (2004). Natural Remedies: Their Origins and Uses. Eds. Taylor and Francis, London. pp.1-169.
- Schonfeld W, Weiland J, Lindig C, Masnyk M, Kabat MM, Kurek A, Wicha J and Repke KR (1985). The lead structure in cardiac glycosides is 5β, 14β-

androstane-3β 14-diol. *Naunyn. Schmiedebergs. Arch. Pharmacol.*, **329**: 414-426.

- Sofowora A (1985). Medicinal plants and traditional medicine in Africa. Wiley, New York.
- Spies T, Praznik W, Hofinger A, Altmann F, Nitsch E and Wutka R (1992). The structure of the fructan sinistrin from *Urginea maritima*. *Carbohydr. Res.*, **235**: 221-230.
- Vega FA, Garcia Jalon, I, Fernandez M and Renedo J (1972). Anthocyanins of red squill, Urginea maritima. Phytochemistry, 11(9): 2896.